

Accumulation and Biotransformation of Mercury
and its Relationship to Selenium after Exposure to
Inorganic Mercury and Methyl Mercury -
A Study on Individuals with Amalgam Fillings,
Dental Personnel, and Monkeys.

by

Magnus Nylander

Department of Environmental Hygiene
and
Institute of Environmental Medicine,
Karolinska Institutet



Stockholm 1990

**Mercury and Selenium Concentrations
and
Their Inter-Relationships in Organs from
Dental Staff and the General Population**

by
Magnus Nylander¹

Jan Weiner²

1. Department of Environmental Hygiene,
Karolinska Institute, P.O. Box 604 00, 104 01
Stockholm, Sweden

2. National Board of Occupational Safety and
Health, 171 84 Solna, Sweden

ABSTRACT

Mercury and selenium concentrations in organs samples from cadavers were determined by radiochemical neutron activation analysis. The relationship between the elements was analysed by linear regression. In pituitary gland samples and one thyroid gland sample from dental staff, occupationally exposed to mercury vapour, accumulation of Se together with Hg at an approximate 1:1 stoichiometric ratio was found. Biological half-times of several years of the accumulated elements were indicated. Accumulation of Se together with Hg at an approximate 1:1 stoichiometric ratio was seen also in renal cortex samples from the general population, i.e. individuals exposed to inorganic mercury from amalgam fillings and organic mercury from food including fish. The possibility of a protective effect of Se against the toxicity of Hg is discussed. The amount of Se that was not associated to Hg, which probably represents the biologically available Se, varied markedly between organs. Decreasing concentrations of biologically available Se with advancing age was seen in pituitary gland samples, but not in other organ samples. Renal cortex samples from three dentists showed low Se relative to Hg. It was suggested that a comparatively large fraction of Hg was bound to other ligands than Se or that the biologically available amount of Se had decreased. The results show the importance of simultaneous analysis of Hg and Se when evaluating organ concentrations of these elements.

Keywords: mercury, selenium, organ concentrations, trace element interactions,

INTRODUCTION

Mercury (Hg) is a non-essential heavy metal the compounds of which differ in metabolism and toxicity. The adverse effects of exposure to elemental mercury vapour (Hg^0) have been known for centuries. After uptake in the lungs Hg^0 is oxidised to mercuric mercury (Hg^{2+}) in the blood and other tissues. In the non-oxidised form it readily passes through biological membranes, e.g. the blood brain barrier and the placenta. The central nervous system and the kidney are considered the critical organs at long time exposure to elemental mercury. The classical intoxication picture, mercurialism, includes tremor, psychic symptoms, notably erethism, and gingivitis.¹ Animal studies suggest that the immune system also might be a critical organ at exposure to inorganic mercury, especially in genetically susceptible individuals.^{2,3}

In the 1960's organic mercury compounds, mainly methyl mercury (MeHg), were demonstrated as widely spread environmental dangers. MeHg is highly lipid soluble and passes readily into all tissues. The central nervous system is considered the critical organ for methyl mercury in man.^{1,4}

As a result of preparing and working with amalgam dental staff is exposed to mercury in the form of vapour and particulates.⁵⁻⁸ This exposure implies a potential risk for deleterious effects.⁹⁻¹³

Recently interest has focused also on the possibility of health effects in the general population due to exposure to inorganic mercury from amalgam restorations.¹⁴⁻¹⁶ Amalgam fillings continuously emit elemental mercury vapour which is absorbed in the lungs, and distributed to various organs.^{8,15,17-19} This mercury makes the predominant contribution to exposure to inorganic mercury in the general population.²⁰

Selenium (Se) is an essential trace element in several species including man. It is an integral part of the enzyme glutathione peroxidase in man and other mammals. It may also have other important functions. At high doses Se becomes toxic, e.g. through intake with food in some geographical areas with high Se content in the soils.²¹ Organ concentrations of minerals and trace elements depend on age. For different organs there have been reports of increasing Se concentrations,^{22,23} but also decreasing Se concentrations²² with advancing age in adults. Plasma Se has been reported to decline markedly after 65 years of age.²⁴

Experimental animal studies show that selenium compounds, both organic and inorganic, protect against the acute renal and intestinal necrosis induced by subcutaneously administered mercuric (Hg^{2+}) chloride.²⁵⁻²⁸ With oral administration the protective effect of selenite against the renal toxicity of HgCl_2 has been demonstrated with exposures up to 20 months.²⁹⁻³¹ Conversely mercury has been demonstrated to protect against the growth retarding effect of toxic doses of selenium in rats and chicks.³⁰⁻³²

After combined administration to rodents, both elements become attached to certain high molecular weight protein fractions in a 1:1 stoichiometric ratio.³³⁻³⁶ The *in vitro* binding of Hg to a single plasma protein in a 1:1 stoichiometric ratio with Se was preceded by the conversion of selenite to Se^{2-} , which may readily react with Hg^{2+} to form HgSe .³⁷ Simultaneous administration also causes changes in the organ distribution of mercury. In most studies a decreased concentration in the kidney and increased concentrations in other organs are reported.^{26,38,39} These changes in distribution are accompanied by increased retention of mercury. The effect on distribution and retention is maximal when the elements are administered in equimolar doses.^{26,35,38-41} Conversely HgCl_2 alters the distribution and retention of selenium.^{35,40}

There are no animal data on the effect of Se treatment on the toxicity of inhaled elemental mercury vapour. A study in mice has shown that except for increased uptake in the lungs Se pretreatment did not markedly change the initial distribution of Hg. However, the retention of mercury in the whole body and several organs was markedly increased. The authors concluded that while injected selenite and mercuric mercury interact strongly in serum and are retained especially in the reticulo-endothelial system, elemental mercury interacts with selenium mainly after oxidation intracellularly.⁴²

In 1975 Kosta et al. reported an approximate 1:1 stoichiometric ratio of Hg and Se at high concentrations in several organs from former mercury miners.⁴³ We also found a very high correlation between high total Hg and Se concentrations in four pituitary glands from dental staff.⁴⁴ The main purpose of the present work was to further elucidate the relationship between mercury and selenium concentrations in several tissues from individuals with varying exposure to mercury vapour.

MATERIAL AND METHODS

Pituitary gland, occipital cortex, renal cortex, abdominal muscle as well as a few samples from thyroid gland were collected at autopsy of 7 dentists and one dental assistant (Table 1). The samples were collected at different pathology departments and at the coroner's office in Stockholm. Pituitary gland, occipital cortex, renal cortex, and abdominal muscle samples, from individuals without known occupational exposure to Hg were also collected (Table 2). All deaths had been sudden and unexpected. The sampling was carried out at the coroner's office in Stockholm.

Two of the individuals with no known occupational exposure to Hg (Nos. 8 and 20, Table 2) had much (>10-fold) higher Hg concentrations than the rest in the pituitary gland. This suggests that occupational exposure may have occurred. They had retired due to old age and with no data on former occupations at the time of sample collection. Later retrieval of data from the National population and housing census of 1980, showed that one of these cases had worked as an electrician. A major

industrial usage of mercury is in electrical equipment,¹ therefore an occupational exposure is not unlikely. For the other individual, no data on former occupations could be obtained.

According to detailed medical records, one dentist (case No.1, Table 1) had not worked for several years due to retirement and incapacitating chronic illness. Another dentist (case No.2) had not worked since retirement, a total of 15 years, according to information from relatives. Detailed data on the remaining cases were not obtained. However, based on the medical records and autopsy reports we concluded that they had not been professionally active during the last months before death. There were no reported signs or symptoms of mercury intoxication in available medical records. However, one of the dentists (Case No. 2) suffered from disturbances of peripheral nerves and was at the time of death investigated for these problems at a neurological clinic.

After subsampling and dissection, analysis of total Hg and Se was carried out in collaboration with the Swedish Environmental Research Institute (IVL) using a radiochemical neutron activation (RNAA) method.⁴⁵⁻⁴⁷ Detailed description and quality control for the mercury analyses have been published earlier.¹⁵ The detection limit is 2 µg/kg for Se. The accuracy of the RNAA method (IVL) to determine Se was established by analyses of standard reference material (SRM) 1577a from the National Bureau of Standards (NBS), with a certified value of 0.71 ± 0.07 mg Se/kg dry weight. The results of three analyses were 0.79, 0.75 and 0.78 mg Se/kg dry weight. A few samples (Cases Nos. 3 and 7, Table 1) were analysed by RNAA at Isotopcentralen, Denmark. The detection limit is 0.2 µg/kg for Hg and 3 µg/kg for Se.⁴⁸ The accuracy was tested through simultaneous analysis of NBS SRM 1577, certified for 16 ± 2 µHg/kg and 1.1 ± 0.07 mg Se/kg, which gave 16.0 and 15.8 µg Hg/kg and 1.1 and 1.1 mg Se/kg.

For the organs with 6 or more samples the relationship between Se, Hg and age was investigated by linear regression analysis⁴⁹, with Se as dependent variable. In all analyses Hg was entered as an independent variable then age was added to the model if this resulted in a significant improvement (Partial F-test $p < 0.10$). Examination of residuals indicated the linear models to be adequate. Based on previous studies it was decided that the alternative to the null hypothesis of no association between Hg and Se is a positive association, therefore significance tests regarding the effect of Hg were performed as one-sided tests. Significance tests regarding the effect of age were performed as two-sided tests. All regression analyses were performed using the SAS statistical package for VAX/VMS.⁵⁰

Mercury and selenium in organs from dental staff

The organ samples from dental staff had considerably higher Hg concentrations than those from the non-occupationally exposed cases. The pituitary glands of the dental staff also had considerably higher Se concentrations than pituitary glands from non-occupationally exposed cases. Significantly higher concentration of Se for dental staff was also seen in occipital cortex (Mann-Whitney test⁵¹), although the magnitude of the difference was less than for pituitary gland (Tables 1 and 2).

Regression analysis of data from pituitary gland with Se concentration as dependent variable demonstrated a strong effect of Hg concentration. The regression coefficient for Hg was 1.0. The intercept (the predicted Se concentration at zero Hg) was 4.6 $\mu\text{mol Se/kg}$, however, this parameter estimate was not statistically different from zero (Figure 1 and Table 3). Addition of age resulted in a borderline significant (Partial F-test $p < 0.11$) improvement, which effect if included in the model would show a negative parameter estimate. The coefficient of determination of the model including also age was, $R^2 = 0.99$.

Regression analysis of data from occipital cortex samples with Se concentration as dependent variable demonstrated an effect of Hg concentration. The regression coefficient for Hg was 1.0, but the confidence interval was wide. No effect of age was seen. The intercept of 2.1 $\mu\text{mol Se/kg}$ showed a significant fraction of Se not associated to Hg (Table 3).

There were too few renal samples (Table 1) to make regression analysis meaningful. The association of Se to Hg generally can not be assessed in single samples, but at the extremely high concentrations of the elements in one thyroid sample (case No. 2) the stoichiometric relationship of approximately 1:1 should be noticed.

Mercury and selenium in organs from individuals without occupational exposure to Hg

In average renal cortex showed the highest Hg concentration. Regression analysis showed a strong relationship between Hg and Se. The regression coefficient for Hg was 1.1. No effect of age was seen. The intercept of 8.5 $\mu\text{mol Se/kg}$ showed a significant fraction of Se not associated to Hg (Table 3, Figure 2).

Regression analysis of data from pituitary gland samples with Se concentration as dependent variable demonstrated a clear effect of both Hg concentration and of age. The regression coefficient for Hg was 1.3. The regression coefficient for age was negative, i.e. decreasing concentrations with advancing age. At a Hg concentration of zero and 58 years of age (average age of cases with pituitary data) the mean Se concentration predicted by this linear model was 5.8 $\mu\text{mol Se/kg}$, thus showing a sig-

Hg concentrations were excluded (see also material and methods) the point estimate of the effect of Hg remained essentially unchanged, but the effect was no longer statistically significant. The negative effect of age did not change and was still statistically significant. These two cases have a good fit to the linear model based on the pituitary glands of the dental staff cases. Only one of these two cases had data on Hg concentrations in other tissues, but it was notable that the mercury concentrations in these tissues were of about the same level as for the other non-occupationally exposed cases.

Regression analysis demonstrated a relationship between Hg and Se in occipital cortex. The regression coefficient for Hg was 4.6, with a wide confidence interval. No effect of age was seen. The intercept of 1.7 $\mu\text{mol Se/kg}$ showed a significant fraction of Se not associated to Hg (Table 3).

Abdominal muscle had the lowest Hg concentrations among the tissues studied. Regression analysis demonstrated an effect of Hg concentration and a borderline significant ($p < 0.10$) effect of age on Se concentration in this tissue. The regression coefficient for Hg was surprisingly high, however, the confidence interval was wide (Table 3 and Figure 3).

Discussion

Dental staff is occupationally exposed to elemental Hg vapour. Concentrations of Hg in tissues from dental staff cases were higher than in corresponding tissues from non-occupationally exposed cases, most prominently so in the pituitary gland (Table 1 and 2). Even though two of the dentists (cases Nos. 1 and 2) had not been occupationally active for several years, they still had extremely high levels of Hg in some organs. This implies biological half-times for the mercury in the order of years. Very long biological half-times of mercury in the human brain have also been indicated in some previous reports.⁵² In a study on human volunteers with single administration of a small dose of radioactively labelled mercury Hursh et al. reported the biological half-time of mercury accumulated; in the kidney region to be approximately 60 days; in the head to be approximately 20 days and in the other gross regions of the body (except lungs) to be in between these figures.⁵³ Thus different fractions of mercury with very large variations in biological half-times appear to exist.

In the non-occupationally exposed individuals the highest concentration of mercury was seen in the kidney. Short term animal studies with exposure to mercury vapour have shown the highest concentrations to accumulate in the kidney.^{42,54-56} In human volunteers the kidney region accumulated the highest levels of radioactive mercury.⁵³ However, in dental staff (Table 1) and mercury retired miners⁴³ the highest Hg concentrations were not seen in the kidney.

Also the levels of Se were increased in pituitary gland of dental staff. Regression analysis indicated that Se accumulated together with Hg at a relationship consistent with a 1:1 stoichiometric ratio in the pituitary gland (Figure 1 and Table 3). Accumulation of Se together with Hg was seen also in occipital cortex, but here the quantitative estimate of the relationship was subject to a larger random variation (Table 3). The data from one thyroid gland, with extremely high concentrations of the elements, indicated accumulation at a 1:1 stoichiometric ratio.

The results are in close agreement with those of Kosta et al.⁴³ who reported accumulation of Se together with Hg at a 1:1 stoichiometric ratio in several organs from five mercury miners, retired since 5-16 years. The relationship between concentrations of mercury in different organs was similar to those in samples from dental staff in the present study, but the levels of Hg and Se were higher. There were no statements regarding health status in the deceased subjects. However, the authors conclusion that the interaction between the elements protected against the toxic properties of these seems to indicate that at least there were no evident signs or symptoms that they related to Hg or Se toxicity.

Ashe et al.⁵⁷ exposed rabbits intermittently (5 days/week, 7 h/day) to high levels (0.9 mg/m³) of mercury vapour for 1-12 weeks. Pathologic changes in brain tissue were seen at organ levels from 1-2 $\mu\text{mol Hg/kg}$ wet weight. Kidney tissue showed pathologic changes at levels slightly below 100 $\mu\text{mol Hg/kg}$. In the heart pathologic changes were seen at slightly lower organ concentrations than in the brain, and in the liver and lungs at slightly higher levels than in the brain. Fukuda et al.⁵⁸ exposed rabbits (4 days/week, 6 h/day) to high levels of mercury vapour (4 mg/m³) for 13 weeks. Tremor and clonus were seen at concentrations of 2-15 $\mu\text{mol Hg/kg}$ wet weight in different parts of the brain. Selenium status was not considered in these studies.

Thus, in several organ samples from dental staff Hg concentrations were in same order of magnitude as those where toxic effects have been seen in animal studies. The relationship between organ levels of mercury and toxic effects in humans is not known.¹

The findings of an association between Hg and Se, with long biological half-time, in several human organs after long term exposure to elemental mercury vapours is in agreement with findings in experimental animal studies with administration of inorganic mercury.^{29,36,40,42} The very high tissue concentrations of Hg that were demonstrated in samples from human organs in the present study and that of Kosta et al. may suggest that this Hg has a comparatively low biological activity. The association of Se to Hg may provide a mechanism of inactivation of Hg and Se. This would be in agreement with data from animal studies where it appears that the formation of protein-Hg-Se complexes or HgSe, which diverts Hg from binding-sites where its toxic effect is normally exerted, is of importance for the protection by Se against the toxicity of Hg and vice versa.^{26,28,30-32,34,59} However, neither in the

cases of the present study nor in those of Kosta et al. can toxic effects of Hg exposure be excluded. Furthermore the high organ concentrations of Hg appear to be at least partly due to an increased retention resulting from the association with Se.

A protective effect might, as proposed by Frost,⁶⁰ imply that the toxic effect of mercury becomes manifest when the amount of Se²⁻ is inadequate to inactivate Hg. A recent paper reported on two Swedish goldsmiths who developed symptoms of mercury intoxication after being unknowingly exposed to mercury vapour during recovery of gold from metal scraps of dental restorations. After cessation of exposure both slowly recovered. Interestingly, during the recovery period, both reported alleviation of symptoms in connection with supplementation of 50 µg organically bound Se daily.⁶¹ This is a moderate amount of Se, but still it is nearly twice the supply in an average Swedish diet,⁶² which is considered suboptimal in Se.⁶³ The possibility of a protective effect of Se against Hg toxicity in long term exposure in humans needs further clarification.

A linear relationship indicating that Se accumulated together with Hg was seen also in the renal cortex of non-occupationally exposed individuals (Figure 2). This relationship was consistent with an accumulation at a 1:1 stoichiometric ratio. In occipital cortex and pituitary gland of non-occupationally exposed individuals a relationship between the elements was also seen, but it was less clear than in the renal cortex with higher mercury concentrations. A significant part of the mercury in human organs, i.e. brain, kidney and pituitary gland has been shown to emanate from amalgam fillings.^{8,15,17-19} The other major source of mercury exposure in the general population is through intake of MeHg with food including fish.¹ In contrast to the occupational exposure in dental staff this exposure continued until death.

A recent study of Japanese forensic cases from the general population also showed a high correlation between inorganic Hg and selenium in renal cortex, but no correlation in cerebrum. Concentrations of mercury in different organs of these cases were markedly higher than for the non-occupationally exposed individuals in the present study, probably resulting from a higher intake of methyl mercury with food including fish in the Japanese population.^{64,65}

A correlation between Hg and Se was seen also in abdominal muscle, but the point estimate of the regression coefficient for Hg suggested the possibility of a relationship different from that seen in the other tissues. There were two cases with high Se concentration in relation to Hg concentration. If these two cases are excluded, a very good fit to a linear model including only Hg is obtained, $R^2=0.81$ (Figure 3). The regression coefficient for Hg becomes 24 (95%-confidence interval [14, 33]). As two cases were excluded this comparatively high slope coefficient may be an artifact.

For occipital cortex and pituitary gland the linear models explaining Se concentra-

occupationally exposed individuals can be compared. The parameter estimates of the intercept for occipital cortex and of the slope coefficient for Hg for pituitary gland are approximately the same in both groups. For the remaining two parameter estimates the random variations are large, but there is no indication of a difference between the groups.

Both in the occupationally exposed and in the non-occupationally exposed individuals, accumulation of selenium together with mercury was clearly seen only in the organs with the highest concentrations of Hg. There are several possible explanations e.g.; at lower concentrations of Hg an interaction may be obscured by natural variations in the Se concentration; an interaction between Hg and Se takes place only above a certain threshold; as only total Hg was analysed the influence of methyl mercury may obscure a relationship between inorganic Hg and Se.

In animal studies the increased retention of Hg after pretreatment with Se was not seen when the doses of inhaled mercury vapour and injected Se were small. One hypothesis forwarded was that at low concentrations Hg is bound to sites with low capacity but higher affinity than that of the proposed Se metabolite (Se^{2-}).⁶⁶

Based on concentration of Hg in the air⁷ and urinary excretion of Hg,⁶⁷⁻⁶⁹ which is a good indicator of ongoing exposure to Hg vapour,^{1,70} the present exposure to mercury in Swedish dentistry has been shown to be moderate. The contribution of occupational exposure to urinary excretion of Hg in dental staff was approximately the same as the contribution from dental amalgam fillings, i.e. in average about 1-3 $\mu\text{g Hg/l}$ or g creatinine from both sources. Urinary excretion of mercury ranged up to approximately 35 $\mu\text{g Hg/l}$ or g creatinine in dental staff and about 15 $\mu\text{g/l}$ or g creatinine in non-occupationally exposed subjects.^{67-69,71} Air concentrations of mercury and urinary mercury excretion in dental staff was lower in public health dentistry than in private dental care.^{7,67,68} Considering the moderate urinary mercury levels in dental staff the very large difference between pituitary Hg concentrations in dental staff and non-occupationally exposed individuals is somewhat surprising. Three dentists had about 200 times higher mercury concentration in the pituitary gland than the median for non-occupationally exposed (Tables 1 and 2).

The highly elevated mercury concentrations in pituitary gland samples from dental staff might be the result of higher exposure earlier in dentistry. Two reports on urinary Hg concentrations in dental staff from 1957⁷² and 1970⁷³ did not, however, indicate a markedly higher exposure previously in Swedish dentistry. These studies were limited and did not present an adequate analytical control, therefore they do not allow a precise assessment of previous exposure.

The female dentist that was only 30 years old at death had worked in public health dentistry and had given birth to two children after graduation as a dentist. Therefore she had only had limited duration of exposure to contemporary levels in dentistry. Still the concentration of Hg in her pituitary gland was more than 10 times higher than the median value of non-occupationally exposed.

Thus there seems to exist factors other than the relation between average levels of exposure that is of importance for the difference in mercury levels in pituitary gland between the two groups. This large difference might partly be the result of an interaction between Hg and Se, that takes place only above a certain threshold, and results in long term accumulation of the elements. Possibly the exposure pattern in dentistry with comparatively low exposure most of the time, but with high exposure peaks, e.g. when preparing amalgam or when drilling in old amalgam fillings^{5,72,74,75} may be of importance for the organ distribution and sub-cellular binding of Hg. The difference between the groups was not as large in occipital cortex where also the interaction between the elements appeared to be less pronounced.

In mammals MeHg is converted to inorganic mercury mainly in the liver, which then is redistributed also to other organs, especially the kidney.^{1,76-79} In several tissues of humans with high oral intake of MeHg a significant fraction of the mercury has been found to be in inorganic form. The kidney usually contained the highest fraction of inorganic Hg.¹ Part of the mercury associated with selenium in the kidney probably emanates from intake of methyl mercury.^{64,65}

Mercuric mercury does not readily pass the blood brain barrier. Significant accumulation of inorganic Hg emanating from intake of methyl mercury therefore requires demethylation in the brain. Recent data from long term exposure of monkeys to MeHg indicated such demethylation.⁸⁰ Further analysis of brain tissue samples from the same animals showed a high correlation between inorganic Hg and Se. However, total Hg (including MeHg) did not show a relationship with Se (unpublished observation). It is not clear if a significant demethylation occurs in the human brain.⁶⁵ In the present study only total Hg was analysed, therefore organic mercury from food may obscure a relationship between inorganic Hg and Se, especially in the brain.

A chemical association or complex formation involving Hg and Se might also inactivate biologically available Se.⁸¹ However, in organs studied by regression analysis the intercept indicated a substantial amount of Se that was not associated to Hg, i.e. probably representing biologically available Se. There was a significant variation in this amount between tissues. If binding to other metals, e.g. cadmium,⁶⁴ were of importance it would most likely have induced variation in the data that could not be explained by the linear models. The very high correlation between Hg and Se in pituitary gland of dental staff and to some extent also in renal cortex of non-occupationally exposed individuals therefore suggests that in the present material such binding of Se to other elements is less important.

For the pituitary glands non-occupationally exposed cases demonstrated decreasing concentrations of biologically available Se with advancing age. This was also suggested by the dental staff cases. The effect of advancing age on Se concentration in different organs of adults is not well known. In the brain Se has been reported to increase,²³ but also to decrease²² with age. In renal cortex it has been reported to increase with age.²² We did not find an effect of age on biologically available Se in

these organs. However, none of the previous studies considered Hg concentrations, therefore age effects in these studies may be secondary to variations in Hg concentration that correlated to age.

Compared to the linear models based on data from the non-occupationally exposed individuals three samples from renal cortex of dental staff all lay below the regression line and outside a 95%-confidence interval for predicted values, i.e. showing lower than expected Se concentrations. This may imply that a comparatively large part of the Hg in the kidney of the dental staff cases was not associated to Se. Such Hg may be inactivated by binding to other protective cell-components, e.g. metallothionein, or it may be associated to binding-sites where it is potentially harmful. In animal studies with administration of mercuric chloride in combination with selenite large amounts of both Hg and Se accumulated in the renal tubule cells.²⁹⁻³¹ The renal tubule is a sensitive indicator of effects of occupational exposure to Hg vapour and effects have been demonstrated after moderate exposure.³²⁻³⁴

Alternatively, the amount of Se that was not associated to Hg was lower than in kidney samples of non-occupationally exposed individuals, implying that the mercury might have lowered the amount of biologically available Se.³¹ However, due to the long exposure free period in at least one of the dental staff cases this alternative would seem less likely.

As noticed above the concentration of Hg in the kidney of dental staff was surprisingly low, displaying only moderately increased levels compared to those of non-occupationally exposed subjects. In dental staff case No. 2 (Table 1), who had not been occupationally exposed for several years, the amount of Se accumulated in the kidney was also low compared to that accumulated in pituitary gland and thyroid gland. As an association between the elements is shown to occur in the kidney of non-occupationally exposed individuals such an association seems likely to occur also in the kidney of occupationally exposed. A possible explanation for the relatively low concentration of the elements in the kidney after occupational exposure is that the biological half-time of Hg and Se that are accumulated together might be shorter in the kidney than in pituitary gland, thyroid gland and brain where extremely long biological half-times have been indicated.

In conclusion, this study demonstrated accumulation of Se together with Hg at a relationship consistent with a 1:1 stoichiometric ratio in organs with comparatively high concentrations of mercury from dental staff and the general population. Long biological half-times of the accumulated elements were indicated. The results show the importance of simultaneous analyses of both Hg and Se when organ concentrations of these elements are evaluated.

1. Clarkson TW, Hurst JB, Sager PR, Syversen ULM. Mercury. In: Clarkson TW, Friberg L, Nordberg GF, Sager PR, eds. *Biological monitoring of toxic metals*. New York: Plenum Press, 1988:199-246.
2. Druet P, Bernard A, Hirsch F et al. Immunologically Mediated Glomerulonephritis Induced by Heavy Metals. *Arch Toxicol* 1982;50:187-94.
3. Hultman P. Effects of inorganic mercury on the murine immune system in vivo. Autoimmunity and systemic immune-complex deposits [Doctoral Thesis]. Linköping: Linköping University, 1989. 168 pp.
4. Berglund F, Berlin M, Birke G, et al. Methyl mercury in fish. A toxicologic-epidemiologic evaluation of risks. Report from an expert group. *Nord Hyg Tidskrift* 1971;suppl 4:1-364.
5. Buchwald H. Exposure of dental workers to mercury. *Am Ind Hyg Assoc J* 1972;33:492-502.
6. Naleway C, Sagaguchi R, Mitchell E, Muller T, Ayer WA, Hefferen JJ. Urinary mercury levels in US dentists, 1975-1983: Review of health assessment program. *J Am Dent Assoc* 1985;111:37-42.
7. Nilsson B, Nilsson B. Mercury in dental practice. I. The working environment of dental personnel and their exposure to mercury vapor. *Swed Dent J* 1986;10:1-4.
8. Nylander M, Friberg L, Eggleston D, Björkman L. Mercury accumulation in tissues from dental staff and controls in relation to exposure. *Swed Dent J* 1989;13:235-43.
9. Cook TA, Yates PO. Fatal mercury intoxication in a dental surgery assistant. *Br Dent J* 1969;127:533-5.
10. Iyer K, Goodgold J, Eberstein A, Berg P. Mercury poisoning in a dentist. *Arch Neurol* 1976;33:788-90.
11. Smith DL. Mental effects of mercury poisoning. *South Med J* 1978;71:904-5.
12. Shapiro IM, Comblath DR, Sumner AJ et al. Neurophysiological and neuropsychological function in mercury-exposed dentists. *Lancet* 1982;i:1147-50.
13. Hryhorczuk DO, Meyers L, Chen G. Treatment of mercury intoxication in a dentist with N-Acetyl-D,L-penicillamine. *J Toxicol Clin Toxicol* 1982;19:401-8.

review of the literature. *Environ Res* 1987;42:257-74.

15. Nylander M, Friberg L, Lind B. Mercury concentrations in the human brain in relation to exposure from dental amalgam fillings. *Swed Dent J* 1987;11:179-87.

16. Weiner JA, Nylander M, Berglund F. Does mercury from amalgam restorations constitute a health hazard? *Sci Total Environ* (in press).

17. Eggleston DW, Nylander M. Correlation of dental amalgam with mercury in brain tissue. *J Prosthet Dent* 1987;58:704-7.

18. Schiele R. Quecksilberabgabe aus Amalgam und Quecksilberablagerung in Organismus und Toxikologische Bewertung. In: Knolle G, ed. *Amalgam - Pro und Contra. Statements - Discussion*. Köln: Deutsche Ärzte-Verlag. 1988:123-131.

19. Drasch G, Schupp I, Günther G. Einfluss von Amalgam-Füllungen auf die Quecksilberkonzentration in der Nierenrinde. In: Anke M, Baumann W, Bräunlich H, Brückner C, Groppe B, Grün M, eds. *Proceedings of the 6th International Trace Element Symposium in Leipzig*. Jena: Der Friedrich Schiller Universität. 1989:1653-1659.

20. Clarkson TW, Friberg L, Hursh JB, Nylander M. The prediction of intake of mercury vapor from amalgams. In: Clarkson TW, Friberg L, Nordberg GF, Sager PR, eds. *Biological Monitoring of Toxic Metals*. New York: Plenum Press, 1988:247-264.

21. Högborg J, Alexander J. Selenium. In: Friberg L, Nordberg GF, Vouk VB, eds. *Handbook on the Toxicology of Metals, 2nd Edition, Volume II: Specific Metals*. Amsterdam: Elsevier Scientific Publishing Company. 1986:482-520.

22. Persigehl M, Schicha H, Kasperek K, Feinendegen LE. Behaviour of trace element concentration in human organs in dependence of age and environment. *J Radioanal Chem* 1977;37:611-5.

23. Duflo H, Maerhaut W, De Reuck J. Regional distribution of potassium, calcium, and six trace elements in normal human brain. *Neurochem Res* 1989;14:1099-112.

24. Bortoli A, Fazzin G, Marchiori M, Mello F, Brugioli R, Martelli F. Plasma selenium in old age as investigated by the effects of selenium supplementation with Se enriched tablets [Abstract]. *J Trace Elem Exp Med* 1989;2:75-.

25. Parizek J, Ostadalova I. The protective effect of small amounts of selenite in sublimate intoxication. *Experientia* 1967;23:142-3.

26. Eybl V, Sykora J, Merl F. Einfluss von Natriumselenite, Natriumtellurit und Natriumsulfit auf Retention und Verteilung von Quecksilber bei Mäusen. *Arch Toxikol* 1969;25:296-305.
27. Magos L, Clarkson TW, Hudson R. Differences in the effects of selenite and biological selenium on the chemical form and distribution of mercury after the simultaneous administration of HgCl₂ and selenium to rats. *J Pharmacol Exp Ther* 1984;228:478-83.
28. Magos L, Clarkson TW, Sparrow S, Hudson AR. Comparison of the protection given by selenite, selenomethionine and biological selenium against the renotoxicity of mercury. *Arch Toxicol* 1987;60:422-6.
29. Groth DH, Stettler L, Mackay G. Interactions of mercury, cadmium, selenium tellurium, arsenic and beryllium. In: Nordberg GF, ed. *Effects and Dose-Response Relationships of Toxic Metals*. Amsterdam: Elsevier Scientific Publishing Company, 1976: 527-543.
30. Carmichael NG, Fowler BA. Effects of combined chronic mercuric chloride and sodium selenite administration in rats: Histological, ultrastructural and X-ray microanalytic studies of liver and kidney. *J Environ Pathol Toxicol* 1980;3:399-412.
31. Lindh U, Johansson E. Protective effect of selenium against mercury toxicity as studied in the rat liver and kidney by nuclear analytical techniques. *Biol Trace Elem Res* 1987;12:109-20.
32. Hill CH. Reversal of selenium toxicity in chick by mercury, copper and cadmium. *J Nutr* 1974;104:593-8.
33. Burk RF, Foster KA, Greenfield PM, Kiker KW. Binding of simultaneously administered inorganic selenium and mercury to a rat plasma protein. *Proc Exp Soc Biol Med* 1974;145:782-5.
34. Chen RW, Whanger PD, Fang SC. Diversion of mercury binding in rat tissues by selenium: A possible mechanism of protection. *Pharmacol Res Comm* 1974;6: 571-9.
35. Fang SC. Interaction of selenium and mercury in the rat. *Chem Biol Interactions* 1977;17:25-40.
36. Naganuma A, Imura N. Properties of mercury and selenium in a high-molecular weight substance in rabbit tissues formed by simultaneous administration. *Pharmacol Biochem Behav* 1981;15:449-54.

37. Gasiewicz TA, Smith JC. Properties of the cadmium and selenium complex formed in rat plasma in vivo and in vitro. *Chem Biol Interactions* 1978;23:171-83.
38. Moffitt AE, Clary JJ. Selenite-induced binding of inorganic mercury in blood and other tissues in the rat. *Res Commun Chem Pathol Pharmacol* 1974;7:593-603.
39. Magos L, Webb M. Differences in distribution and excretion of selenium and cadmium or mercury after their simultaneous administration subcutaneously in equimolar doses. *Arch Toxicol* 1976;36:63-9.
40. Hansen JC, Kristensen P. Organ clearance of SeO_2 and HgCl_2 administered separately and simultaneously to mice. *Toxicology* 1979;15:1-7.
41. Kristensen P, Hansen JC. Wholebody elimination of SeO_2 and HgCl_2 administered separately and simultaneously to mice. *Toxicology* 1979;12:101-9.
42. Khayat A, Dencker L. Interactions between selenium and mercury in mice: Marked retention in the lung after inhalation of metallic mercury. *Chem Biol Interactions* 1983;46:283-98.
43. Kosta L, Byrne AR, Zelenko V. Correlation between selenium and mercury in man following exposure to inorganic mercury. *Nature* 1975;254:238-9.
44. Nylander M, Weiner J. Relation between mercury and selenium in pituitary glands of dental staff. *Br J Ind Med* 1989;46:751-2.
45. Sjöstrand B. Determination of mercury and arsenic in biological and organic material by activation analysis. *Anal Chem* 1964;36:814-.
46. Ljunggren K, Johnels AG, Olsson M, Otterlind G, Sjöstrand B, Westermarck T. Activation analysis of mercury in water and aquatic ecosystems. Conf. IAEA-SM-142 a/22. 1971.
47. Dusan B, Krjstiansson S, Lundberg Å, Skärdin I-L, Werner J, Österdahl G. Routine analyses of mercury. A comparative study between two methods; neutron activation and flameless atomic absorption. Report B 758. The Swedish Environmental Research Institute, Stockholm, Sweden, 1984.
48. Drabæk I, Carlsen V, Just L. Routine determination of mercury, arsenic and selenium by radiochemical neutron activation analysis. *J Radioanal Nucl Chem Lett* 1986;4:249-60.
49. Draper NR, Smith H. Applied regression analysis. 2nd ed. New York: Wiley & Sons. 1981.

50. SAS User's Guide: Statistics, Version 5 Edition. Cary, North Carolina: SAS Institute Inc., 1985.
51. Snedecor GW, Cochran WG. Statistical methods, Seventh Edition. Ames, Iowa: The Iowa State University Press, 1987:144-5.
52. Cavanagh JB. Long term persistence of mercury in the brain [Editorial]. *Br J Ind Med* 1988;45:649-51.
53. Hursh JB, Clarkson TW, Cherian MG, Vostal JJ, Maille RV. Clearance of mercury (Hg-197, Hg-203) vapor inhaled by human subjects. *Arch Environ Health* 1976;31:302-9.
54. Berlin M, Ullberg S. Accumulation and retention of mercury in the mouse. *Arch Environ Health* 1963;6:589-601.
55. Rothstein A, Hayes A. The turnover of mercury in rats exposed repeatedly to inhalation of vapor. *Health Phys* 1964;10:1099-113.
56. Khayat A, Dencker L. Whole body and liver distribution of inhaled mercury vapor in the mouse: Influence of ethanol and aminotriazole pretreatment. *J Appl Toxicol* 1983;3:66-74.
57. Ashe WF, Largent EJ, Dutra FR, Hubbard DM, Blackstone M. Behaviour of mercury in the animal organism following inhalation. *A M A Arch Industr Hyg Occup Med* 1953;7:19-45.
58. Fukuda K. Metallic mercury induced tremor in rabbits and mercury content of the central nervous system. *Br J Ind Med* 1971;28:308-11.
59. Christensen M, Rungby J, Mogensen SC. Effects of selenium on toxicity and ultrastructural localization of mercury in cultured murine macrophages. *Toxicol Letters* 1989;47:259-70.
60. Frost DV. The two faces of selenium - can selenofobia be cured. *CRC Crit Rev Toxicol* 1972;1:467-514.
61. Blomqvist AM, Ahlborg G, Ulander A. Mercury intoxication in goldsmiths after recovery of dental gold? *Läkartidningen* 1989;86:2782-3 (In Swedish).
62. Bruce Å. Medical aspects on selenium. *Kungliga Skogs- och Lantbruks-akademins Tidskrift* 1984;123:267-71.
63. National Research Council - Committee on Dietary Allowances. Recommended Dietary Allowances, 9th ed. Washington DC: Natl. Acad. Sci 1980

relationship between the concentrations of some elements in the organs of Japanese with special reference to selenium-heavy metal relationships. *Sci Total Environ* 1990;91:127-40.

65. Matsuo N, Suzuki T, Akagi H. Mercury concentration in organs of contemporary Japanese. *Arch Environ Health* 1989;44:298-303.

66. Khayat A. Disposition of metallic mercury vapor and mercuric chloride in adult and fetal tissues: influence of pretreatment with ethyl alcohol, aminotriazole, selenium, and tellurium [Doctoral thesis]. Uppsala: Uppsala University, 1985, 47 pp.

67. Nilsson B, Nilsson B. Mercury in dental practice. II. Urinary mercury excretion in dental personnel. *Swed Dent J* 1986;10:221-32.

68. Skare S, Bergström T, Engqvist A, Weiner JA. Mercury exposure of dentists and dental nurses assigned to different origins. *Scand J Work Environ Health* (in press).

69. Åkesson I, Schütz A, Attewell R, Skerfving S, Glantz PO. Status of mercury and selenium tatus in dental personnel - impact of amalgam work and own fillings. *Arch Environ Health* (in press).

70. Roels H, Abdeladim S, Ceulemans E, Lauwerys R. Relationships between the concentration of mercury in air and in blood or urine in workers exposed to mercury vapour. *Ann Occup Hyg* 1987;31:135-45.

71. Langworth S, Elinder C-G, Åkesson A. Mercury exposure from dental fillings. I. Mercury concentrations in blood and urine. *Swed Dent J* 1988;12:69-70.

72. Frykholm KO. Mercury from dental amalgam its toxic and allergic effects and some comments on occupational hygiene. *Acta Odontol Scand* 1957;15 (suppl. 22):1-108.

73. Frykholm KO. Exposure of dental personnel to mercury during work. *Swed Dent J* 1970;63:763-72.

74. Reinhardt JW, Chan KC, Schulein TM. Mercury vaporization during amalgam removal. *J Prosthet Dent* 1983;50:62-4.

75. Richards JM, Warren PJ. Mercury vapour released during the removal of old amalgam restorations. *Br Dent J* 1985;155:231-2.

76. Koeman JH, Peeters WHM, Koustaal-Hol CHM, Tijoe PS, de Goeij JJM. Mercury-Selenium correlations in marine mammals. *Nature* 1973;245:385-6.

77. Norseth T, Clarkson TW. Studies on the biotransformation of ^{203}Hg labeled methyl mercury chloride. *Arch Environ Health* 1970;21:717-27.
78. Syversen TLM. Biotransformation of Hg-203 labelled methyl mercuric chloride in rat brain measured by specific determination of Hg^{2+} . *Acta Pharmacol Toxicol* 1974;35:277-83.
79. Yamamoto R, Suzuki T, Satoh H, Kawai K. Generation and dose as modifying factors of inorganic mercury accumulation in brain, liver and kidneys of rats fed methylmercury. *Environ Res* 1986;41:309-18.
80. Lind B, Friberg L, Nylander M. Preliminary studies on methylmercury biotransformation and clearance in the brain of primates: II. Demethylation of mercury in brain. *J Trace Elem Exp Med* 1988;1:49-56.
81. Wada O, Yamaguchi T, Ono M, Nagahashi M, Morimura T. Inhibitory effect of mercury on kidney glutathione peroxidase and its prevention by selenium. *Environ Res* 1976;12:75-80.
82. Roels H, Gennart J-P, Lauwerys R, Buchet J-P, Malchaire J, Bernard A. Surveillance of workers exposed to mercury vapour: validation of previously proposed biological threshold limit value for mercury concentration in urine. *Am J Ind Med* 1985;7:45-71.
83. Barregård L, Hultberg B, Schütz A, Sällsten G. Enzymeuria in workers exposed to inorganic mercury. *Int Arch Occup Environ Health* 1988;61:65-9.
84. Verschoor MA, Herber RFM, Zielhuis RJ. Urinary mercury levels and early changes in kidney function in dentists and dental assistants. *Community Dent Oral Epidemiol* 1988;16:148-52.

Table 1. Mercury and selenium concentrations
from dental personnel*

| Case No. | Sex | Age years | Pituitary gland | | Occipital cortex |
|----------|-----|-----------|--------------------------|-----|--------------------------|
| | | | Hg $\mu\text{mol/kg}$ | Se | Hg $\mu\text{mol/kg}$ |
| 1 | M | 80 | 20 | 23 | 1.43 |
| 2 | M | 82 | 18 | 22 | 0.42 |
| 3 | M | 60 | 28 | 36 | 0.08 |
| 4 | M | 50 | 1.7 | 9 | 0.19 |
| 5 | F | 30 | 1.5 | - | 0.07 |
| 6 | M | 61 | 0.7 | - | 0.07 |
| 7 | M | 60 | 2.2 | 4.7 | 0.25 |
| 8 | F | 67 | 6.4 | 12 | 0.09 |

*). Cases Nos. 1 to 7 dentists and case N

Medical diagnoses of possible significant concentrations:

case 1: Parkinson's disease and diabetes

case 4: Glomerulonephritis.

case 6: Diabetes mellitus.

Table 2. Mercury and selenium concentration in tissue samples from individuals without occupational exposure to mercury

| Case No. | Sex | Age years | Pituitary gland | | Occipital cortex | | Renal cortex | | Abdominal muscle | |
|----------|-----|-----------|-----------------------|------|-----------------------|-------|-----------------------|------|-----------------------|-------|
| | | | Hg $\mu\text{mol/kg}$ | Se | Hg $\mu\text{mol/kg}$ | Se | Hg $\mu\text{mol/kg}$ | Se | Hg $\mu\text{mol/kg}$ | Se |
| 1 | M | 24 | 0.14 | 11 | 0.050 | 1.90 | 1.63 | 11.1 | - | - |
| 2 | M | 48 | 0.08 | 08 | 0.012 | 1.43 | 0.16 | 8.4 | 0.005 | 1.05 |
| 3 | M | 29 | 0.09 | 07 | 0.032 | 1.82 | 1.27 | 11.4 | 0.011 | 2.38 |
| 4 | M | 63 | 0.17 | 05 | 0.051 | 1.68 | 0.24 | 7.5 | 0.024 | 1.28 |
| 5 | M | 79 | 0.06 | 05 | 0.027 | 1.76 | 0.15 | 8.2 | 0.009 | 1.05 |
| 6 | M | 73 | 0.08 | 07 | 0.020 | 2.00 | 0.11 | 9.6 | 0.010 | 1.27 |
| 7 | M | 80 | - | - | 0.060 | 2.35 | - | - | 0.024 | 1.77 |
| 8 | M | 71 | 3.88 | 14 | 0.114 | 1.96 | 3.79 | 12.8 | 0.047 | 2.01 |
| 9 | M | 74 | 0.08 | 06 | 0.046 | 2.00 | 0.29 | 8.2 | 0.022 | 1.52 |
| 10 | M | 40 | 0.15 | 07 | 0.053 | 2.00 | 1.57 | 9.2 | 0.017 | 1.39 |
| 11 | M | 67 | 0.20 | 06 | 0.061 | 1.81 | 0.52 | 9.7 | 0.024 | 1.53 |
| 12 | M | 16 | 0.19 | 08 | 0.037 | 1.77 | 4.04 | 12.4 | 0.004 | 1.84 |
| 13 | F | 30 | 0.38 | 08 | 0.079 | 1.78 | 2.59 | 11.9 | 0.027 | 1.66 |
| 14 | M | 30 | - | - | 0.036 | 1.84 | - | - | - | - |
| 15 | M | 52 | - | - | 0.098 | 2.08 | - | - | - | - |
| 16 | M | 76 | 0.08 | 04 | 0.049 | 1.92 | - | - | - | - |
| 17 | F | 56 | 0.14 | 05 | - | - | - | - | - | - |
| 18 | M | 61 | 0.03 | 05 | - | - | - | - | - | - |
| 19 | M | 88 | 0.11 | 04 | - | - | - | - | - | - |
| 20 | M | 70 | 5.83 | 11 | - | - | - | - | - | - |
| 21 | M | 71 | 0.05 | 06 | - | - | - | - | - | - |
| 22 | M | 47 | 0.05 | 05 | - | - | - | - | - | - |
| 23 | F | 75 | 0.19 | 03 | - | - | - | - | - | - |
| 24 | M | 63 | 0.04 | 04 | 0.081 | 2.57 | - | - | - | - |
| n | | | 24 | 21 | 17 | 17 | 12 | 12 | 12 | 12 |
| Min | | | 16 | 0.03 | 03 | 0.012 | 1.43 | 0.11 | 7.5 | 0.004 |
| Max | | | 88 | 5.83 | 14 | 0.114 | 2.57 | 4.04 | 12.8 | 0.047 |
| Median | | | 63 | 0.11 | 5.7 | 0.050 | 1.90 | 0.89 | 9.7 | 0.020 |
| Average | | | 58 | 01 | 07 | 0.053 | 1.92 | 1.36 | 10.0 | 0.019 |
| SD | | | 20 | 01 | 03 | 0.027 | 0.26 | 1.42 | 1.7 | 0.012 |

Table 3. Results of linear regression analysis with selenium concentration (umol/kg wet weight) in different tissues as dependent variable*.

| tissue | n | independent variable of parameter | point estimate | 95%-confidence interval | cumulative R ² |
|--|----|-----------------------------------|--------------------|-------------------------|---------------------------|
| <u>Dental staff</u> | | | | | |
| pituitary gland | 6 | intercept ⁺ | 4.6 | [-0.1, 9.2] | 0.96 |
| | | Hg | 1.0 ^a | [0.8, 1.3] | |
| occipital cortex | 7 | intercept | 2.1 ^a | [1.6, 2.7] | 0.57 |
| | | Hg | 1.0 ^a | [0.01, 1.9] | |
| <u>Non-occupationally exposed</u> | | | | | |
| renal cortex | 12 | intercept | 8.5 ^a | [7.7, 9.3] | 0.77 |
| | | Hg | 1.1 ^a | [0.7, 1.6] | |
| pituitary gland | 21 | intercept | 5.8 ^a | [5.0, 6.7] | 0.43 |
| | | Hg | 1.3 ^a | [0.8, 1.9] | |
| | | age | -0.06 ^b | [-0.1, -0.02] | |
| occipital cortex | 17 | intercept | 1.7 ^a | [1.4, 2.1] | 0.24 |
| | | Hg | 4.6 ^c | [-1.8, 10.6] | |
| abdominal muscle | 12 | intercept | 1.2 ^a | [0.8, 1.7] | 0.10 |
| | | Hg | 17.5 ^c | [-4, 39] | |
| | | age | -0.01 | [-0.02, 0.001] | |
| *) Model: [Se] _i = + β ⁰ [Hg] _i + β ¹ age _i + ε _i , ε _i ~ N(0.) | | | | | |

In all analyses the concentration of Hg (umol/kg wet weight) was entered as a dependent variable, if significant improvement (Partial F-test $p < 0.10$) of the model was achieved, age was also included in the model. For the analyses where age (yr) was included it was scaled to actual age minus average age of cases with available samples, i.e. 52 years for non-occupationally exposed subjects with pituitary gland data and 56 years for non-occupationally exposed subjects with abdominal muscle data. Significance test regarding the parameter estimate for the effect of Hg were performed as one-sided T-tests and regarding the effect of age as two-sided T-tests (footnotes a, b and c).

a) $p < 0.001$ b) $p < 0.01$ c) $p < 0.05$

+) The parameter in the model equation, i.e. the predicted Se concentration when all independent variables are zero.

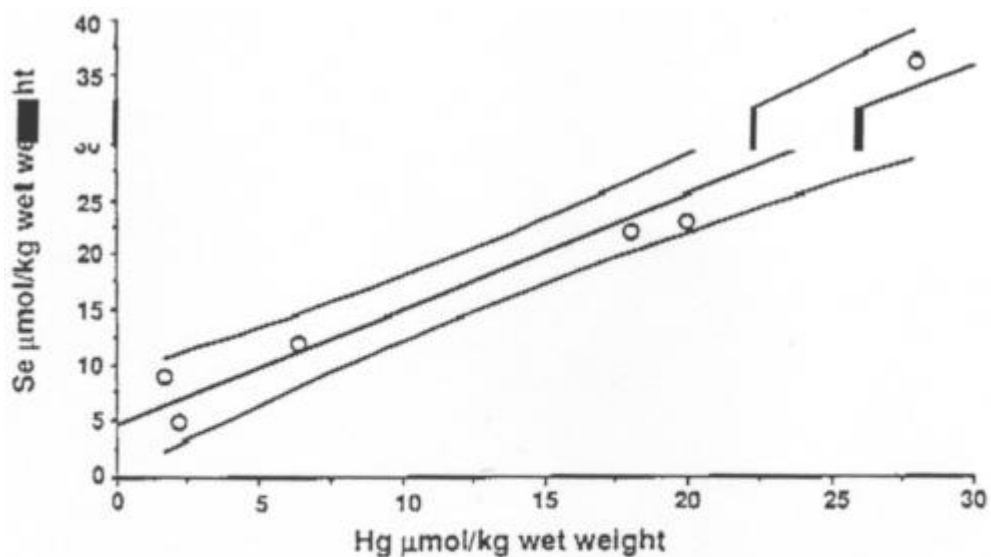


Figure 1. Mercury (Hg) and selenium (Se) concentrations ($\mu\text{mol/kg wet weight}$) in pituitary gland samples from dental personnel. Least squares regression line and 95%-confidence limits for the predicted means.

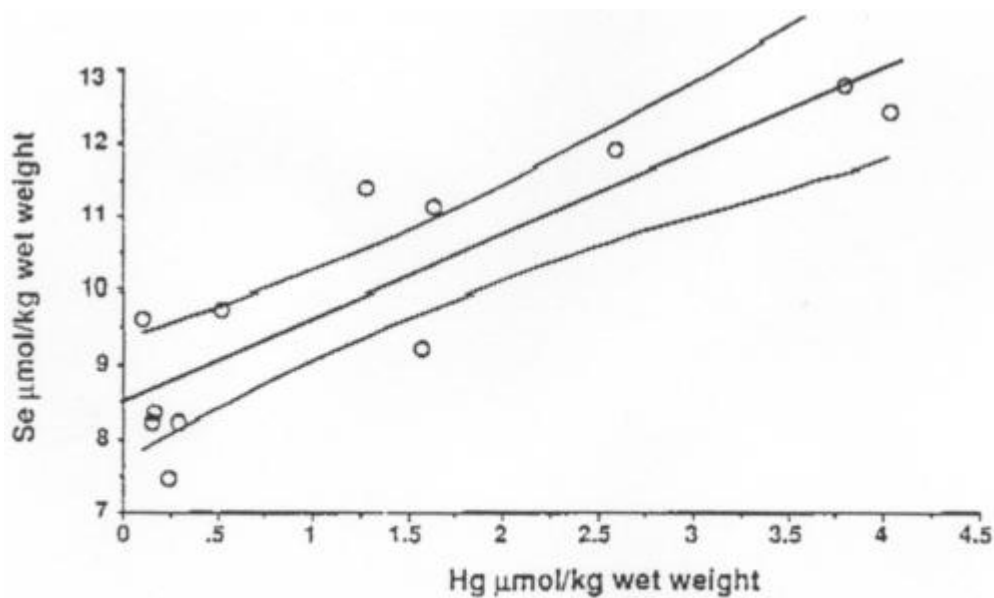


Figure 2. Mercury (Hg) and selenium (Se) concentrations ($\mu\text{mol/kg wet weight}$) in renal cortex samples from non-occupationally exposed individuals. Least squares regression line, 95%-confidence limits for the predicted means.

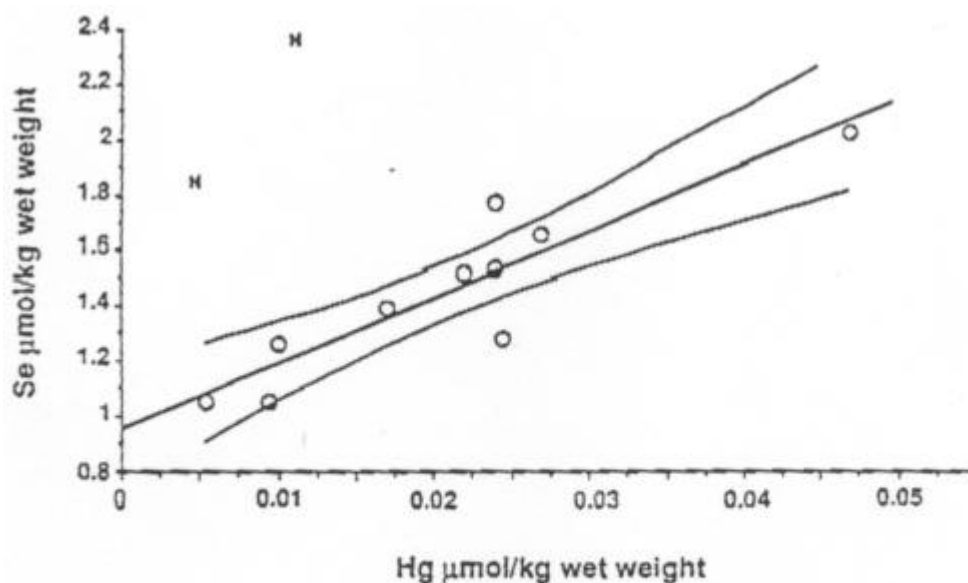


Figure 3. Mercury (Hg) and selenium (Se) concentrations ($\mu\text{mol/kg wet weight}$) in abdominal muscle samples from non-occupationally exposed individuals. Least squares regression line and 95%-confidence limits for the predicted means. Data from two cases are included in the figure (x), but not in the calculation of the regression line